



REVIEW ARTICLE

Open Access



# Soil microbial carbon use efficiency and the constraints

Run Dang<sup>1,2</sup>, Jian Liu<sup>3\*</sup>, Eric Lichtfouse<sup>4</sup>, Lifeng Zhou<sup>1,2</sup>, Meng Zhou<sup>5</sup> and Leilei Xiao<sup>2</sup>

## Abstract

**Background** Microbial contributions to soil organic carbon formation have received increasing attention, and microbial carbon use efficiency is positively correlated with soil organic carbon storage.

**Mainbody** This work reviews the impact on microbial carbon use efficiency from six constraints, including plant community composition and diversity, soil pH, substrate quality, nutrient availability and stoichiometric ratios, soil texture and aggregates, water and thermal constraints, and external nutrient inputs. In general, the response of microbial carbon use efficiency showed large uncertainty to above constraints, including positive-, negative-, or non-correlation. However, some factors are biased, more likely to promote or inhibit carbon use efficiency. For example, external nutrient input (N, P, K, Ca) tended to promote carbon use efficiency, while climate warming showed more negative influence.

**Conclusion** Further, overwhelming works focused on single constraint, we suggest the importance to consider the synergistic influence of multiple environmental variables on microbial carbon use efficiency, special for the regulation mechanism of biological-environmental interactions.

**Keywords** Microbial carbon use efficiency, Environmental constraints, Soil pH, Climate warming, Stoichiometric ratio

## Introduction

Loss of soil organic carbon (SOC) is known to accelerate climate warming, whereas store of more organic carbon into soils can help mitigate climate change. Microbial carbon use efficiency (CUE) is defined as the ratio of carbon consumed by microorganisms for their own growth to the sum of carbon consumed for both growth and respiration (Tao et al. 2023; Fig. 1), which is related to both carbon fixation and loss. Increasing studies propose that minor changes in soil microbial CUE have significant impacts on soil carbon storage and gas emissions (Domeignoz-Horta et al. 2020; García-Palacios et al. 2021; Tao et al. 2023). For example, a high soil microbial CUE implies more accumulation of microbial by-products and residues, which can increase soil carbon storage (Liang et al. 2017, 2019; Sokol et al. 2022). Theoretical analysis and empirical observations showed CUE over

\*Correspondence:

Jian Liu

liujian@dzu.edu.cn

<sup>1</sup>School of Geography and Environment, Liaocheng University, Liaocheng 252059, PR China

<sup>2</sup>CAS Key Laboratory of Coastal Environmental Processes and Ecological Remediation, Yantai Institute of Coastal Zone Research, Chinese Academy of Sciences, Yantai 264003, PR China

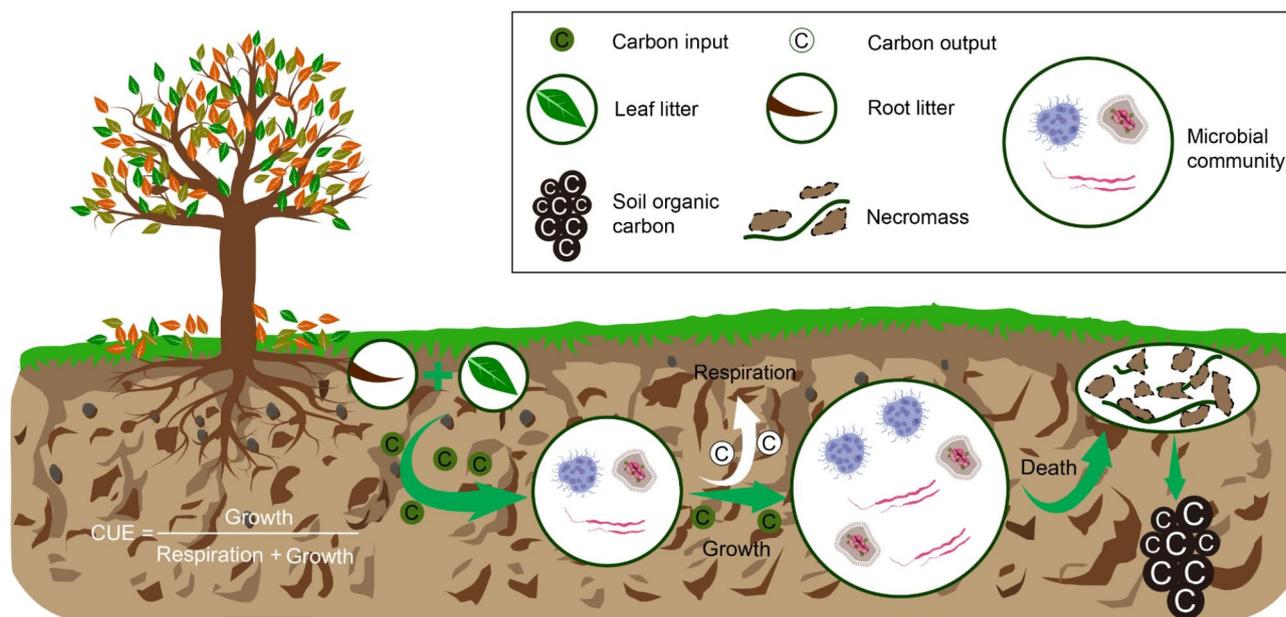
<sup>3</sup>Shandong Key Laboratory of Biophysics, Institute of Biophysics, Dezhou University, No. 566 University Rd. West, Dezhou 253023, PR China

<sup>4</sup>State Key Laboratory of Multiphase Flow in Power Engineering, International Research Center for Renewable Energy, Xi'an Jiaotong University, Xi'an 710049, China

<sup>5</sup>State Key Laboratory of Black Soils Conservation and Utilization, Northeast Institute of Geography and Agroecology, Chinese Academy of Sciences, Harbin 150081, PR China



© The Author(s) 2024. **Open Access** This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit <http://creativecommons.org/licenses/by/4.0/>.



**Fig. 1** Soil microbial carbon use efficiency. Plants primarily input carbon into the soil through root exudates and leaf litter. This litter serves as a carbon source for soil microorganisms, with a portion of the carbon released into the atmosphere through respiratory metabolism. Another portion is assimilated into microbial biomass. Upon death, these microorganisms form microbial residues, which contribute to the accumulation of soil organic carbon. Only a small amount of carbon is consumed for the secretion of extracellular enzymes and metabolic products (negligible). Therefore, microbial carbon use efficiency refers to the ratio of carbon consumed by microorganisms for their own growth to the sum of carbon consumed for both growth and respiration

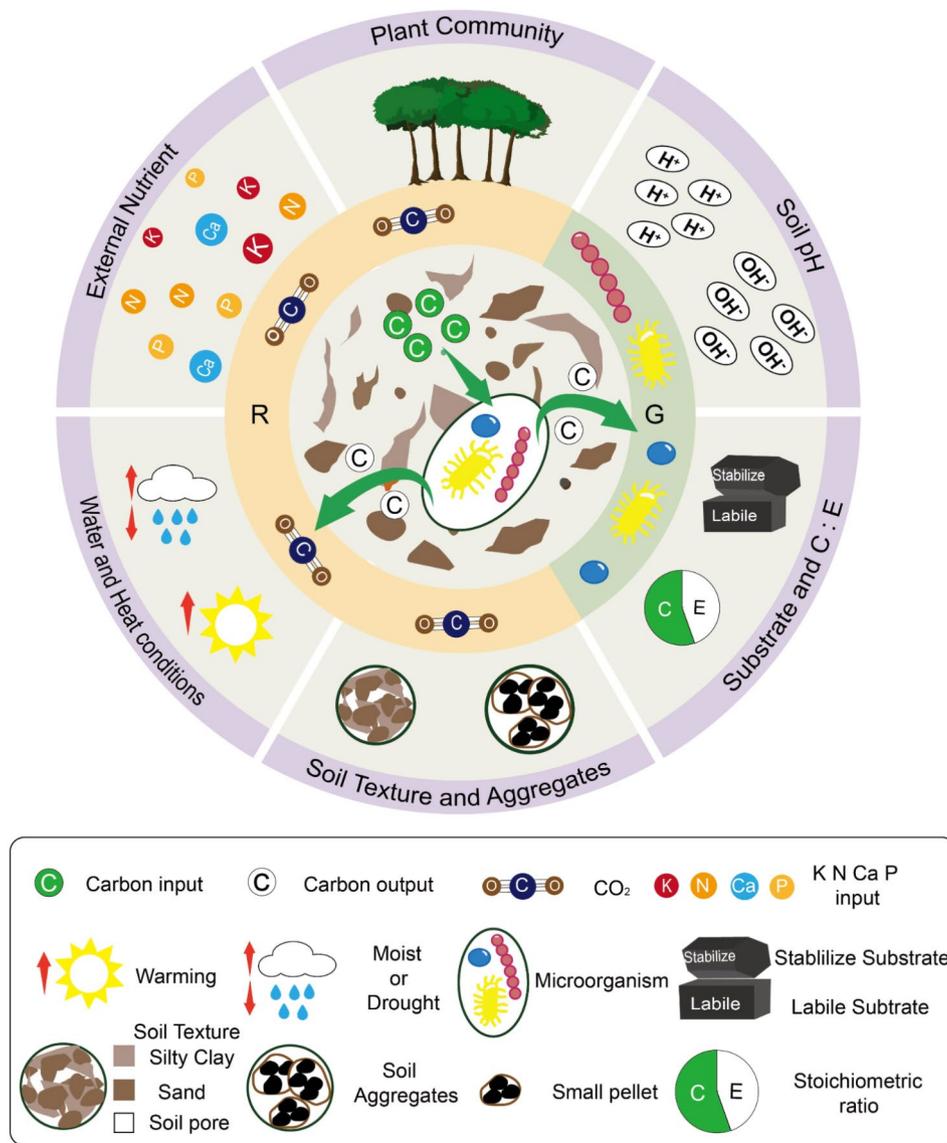
a wide range of field conditions converges around 0.30 (Sinsabaugh et al. 2013). Changes in the abiotic and biotic factors, however, are acknowledged to have a significant impact on soil microbial CUE (Duan et al. 2023; Wang et al. 2023a; Xu et al. 2024).

Up to date, the limited reviews on microbial CUE mainly focused on the methodology, comparing the adaptation conditions and differences of various methods (Manzoni et al. 2012; Frey et al. 2013). For example, each method has its own set of benefits, drawbacks, and application range (Adingo et al. 2021). Some review paper also paid attention to the impact of a certain factor, such as warming (Zhang et al. 2022a), changes in plant types (Zhang et al. 2023a), and biological interactions in soil (Iven et al. 2023). However, the effects of the focused factors on microbial CUE are often multifaceted and unpredictable. A comprehensive analysis of multiple environmental factors remains scarce (Adingo et al. 2021). Here we emphatically discussed on the influences of abiotic and biotic factors on microbial CUE (Fig. 2). It is aimed to provide a comprehensive understanding of the response of microbial CUE to a changing environment, and highlights the focus to improve the prediction of microbial CUE.

### Plant community composition and diversity

Currently, numerous studies have confirmed that plant species diversity significantly impacts the composition, activity, and biomass of soil microbial communities (Lu

and Scheu 2021; Gottschall et al. 2022; Chen and Hu 2024). Plants shape soil conditions (water content, nutrient input) through litter and root inputs, which can promote microbial metabolism and potentially increase CUE (Iven et al. 2023). It indicates a positive correlation between plant community construction and soil CUE. The existence of plants has increased soil water content, thereby enhancing substrate diffusion and improving microbial CUE (Alvarez et al. 2017; Manzoni et al. 2012; Domeignoz-Horta et al. 2020). Higher plant species diversity and primary production increase the input of plant-derived carbon and nutrient content into the soil (Mori et al. 2020; Peng and Chen 2021). By this means, the resource supply for soil microbial communities is enhanced, promoting faster microbial growth, turnover, and higher CUE (Lange et al. 2015; Craig et al. 2022; Xiong et al. 2023). Duan et al. (2023) found that, with increasing tree species diversity and substrate availability, microbial CUE was also increased. The primary succession of vegetation is also accompanied by an increase in community richness. A study in the Hailuoguo glacier retreat area revealed that, with the succession of vegetation, the ratio of oligotrophic to autotrophic microorganisms increased, and CUE gradually increased (Ma et al. 2023a). Moreover, the quantity and type of compounds secreted by plants are specific to plant species (Manzoni et al. 2012). Plants can provide energy sources for microorganisms by secreting labile carbon compounds (Hartmann et al. 2009), which may increase the CUE



**Fig. 2** Response of Soil Microbial Carbon Use Efficiency (CUE) to Different Factors. Microorganisms absorb carbon from the soil, with part of it used for growth(G) and the other part for respiration(R). The inner circle represents the proportion of carbon used for growth in the total carbon absorbed by microorganisms, which is called microbial carbon use efficiency (CUE). Six key factors are covered, starting from the top and proceeding clockwise: plant community composition and diversity; soil pH; substrate quality, nutrient availability and stoichiometric ratios (C: E represents the ratio of carbon (C) content to essential element (E) content in the substrate); soil texture (sand, loam, clay) and aggregates; water and thermal condition limitations (climate warming, drought, and humidity); external nutrient inputs (K, N, Ca, P)

of free-living microorganisms under carbon limitation. Therefore, in general, diverse plant systems have higher microbial biomass and lower respiration rates compared to monocultures (Anderson and Domsch 2010).

Moreover, plants can compete with microorganisms and affect microbial CUE. For example, the abundance of nitrate-reducing bacteria decreases with changes in plant nitrogen use efficiency (Moreau et al. 2015). Through indirect competition, plants may select the proliferation of fast-growing microbes or select for microbes with high investments in resource acquisition. Thus, the community-level CUE is reduced, but such an effect

may additionally depend on the plant species involved and environmental context of the rhizosphere (Iven et al. 2023). Further, some plants even produce secondary metabolites (such as jasmonic acid and salicylic acid) (Watson et al. 2015) or utilize toxic compounds (such as indole glucosinolates) (Anthony et al. 2020) to inhibit related microorganisms, potentially reducing CUE. Contrary to the above view, Prommer et al. (2020) found in a diversity experiment in temperate grasslands that plant species richness had no significant effect on microbial CUE. Cascading effects of biotic interactions are widely known to introduce apparent stochasticity to microbial

communities, which may be difficult to predict (Powell et al., 2016). When examining the interactions between microorganisms and plants, it's important to analyze specific conditions in detail.

#### Soil pH value

The soil pH value is a major factor influencing microbial CUE (Wu et al. 2023, 2024; Liu et al. 2024). Two mechanisms are now proposed influencing the microbial community composition and activity. Firstly, under low pH conditions, bacterial growth is suppressed, leading to decrease organic matter availability (Silva-Sánchez et al. 2019). Less resources resulted in a decreased CUE, indicating a close correlation between pH and CUE (Silva-Sánchez et al. 2019). Secondly, pH impacts microbial activity through cellular stress. For example, under low soil pH conditions, the solubility of toxic metals such as  $Al^{3+}$  increases, causing cellular stress and subsequently reducing microbial CUE (Wang et al. 2023a; Jones et al. 2019). Additionally, microorganisms in low pH environments may need to expend more energy to maintain pH balance, reducing the energy available for growth and thus decreasing microbial CUE (Wu et al. 2023). However, it was also found that the relationship between CUE and soil pH is not a simple positive linear one. With lime addition to agricultural soil, the overall relationship between CUE and pH followed a U-shaped (i.e., quadratic) curve (Schroeder et al. 2024). This suggested that CUE may reach its highest levels under acidic or alkaline pH conditions and be lowest under near-neutral soil conditions. Sinsabaugh et al. (2016) discovered a significant CUE minimum at a pH of 5.4 in a meta-analysis of global soils, which attributed to changes in the bacterial to fungal ratio.

#### Substrate quality, nutrient availability, and stoichiometric ratios

Numerous studies have demonstrated that the quality of carbon substrates, nutrient availability, and stoichiometric ratios regulate microbial CUE (Sinsabaugh et al. 2013; Mooshammer et al. 2014; Li et al. 2021). Complex organic compounds typically have lower carbon substrate quality due to their diverse types of chemical bonds and complex structures, requiring more enzymes for decomposition (Sinsabaugh et al. 2013). The incorporation of such low-quality carbon compounds into microbial biomass leads to microorganisms allocating more energy for enzyme production, reducing the energy available for growth and thus decreasing CUE values (Allison 2014). Such as highly oxidized chemicals like oxalic acid are to be integrated into biomass (Hervé et al. 2016). They necessarily consume significant amounts of reducing power (NADH), while generating a relatively minuscule amount of energy compared to glucose (Hervé et al. 2016). As a

result, on oxalic acid or phenolic substances, the CUE of soil microbial communities is significantly smaller than on glucose (Frey et al. 2013). The degree of C reduction of the substrate ( $\gamma_S$ ) is another important factor that affects the CUE of soil microorganisms. The  $\gamma_S$  of the main substrates utilized by microorganisms is usually within the range of 3–5, which is equivalent to the C reduction degree ( $\gamma_B \approx 4.2$ ) of soil microorganisms (Roels et al. 1980). When the  $\gamma_S$  of the substrate is less than 4.2, the microbial CUE is mainly limited by the reduction degree from the substrate. When the  $\gamma_S$  of the substrate is more than 4.2, the microbial CUE is higher (Roels et al., 1980; Gommers et al. 1988). It is worth noting that the responses of different microbial communities to substrate carbon quality were different. Compared with bacteria exposed to a carbon-rich environment, bacteria exposed to a carbon-limited environment can metabolize a wider range of substrates (von Stockar et al., 2013).

Substrate quality have a significant relative to soil depth. Since the surface soil receives more unstable carbon from surface litter and roots compared to deeper soil layers, this carbon is more readily available for efficient utilization (Zhang et al. 2023b). Deeper soil layers dominated by complex organic compounds lack readily available organic carbon in the form of plant debris and rhizodeposits. This suggesting that CUE may decrease with increasing soil depth. On one hand, under such conditions, it may reduce the decomposition of organic carbon in deeper soil layers (Agren and Bosatta 1987). On the other hand, as soil depth increases, the availability and quality of soil C substrates decrease. As organic carbon concentrations decrease, the benefits of organic carbon decomposition for microorganisms become smaller. The effect of soil depth on microbial CUE may be amplified by changes in organic matter quality, especially for compounds that require extensive enzymatic reactions for degradation (Agren and Bosatta 1987). Even if the microbial community possesses all the enzymes decomposing organic compounds, microorganisms must invest a significant amount of C and N in producing extracellular enzymes, thereby reducing microbial CUE. (Agren and Bosatta 1987; Manzoni and Porporato 2009). Zhang et al. (2023b) discovered that CUE decreased as soil depth in alpine grasslands on the Tibetan Plateau, microbial CUE showed significant negative correlation with microbial biomass-specific hydrolytic enzyme activity.

Under nutrient-limited conditions, microbial CUE decreases accordingly. Microorganisms adapt to nutrient-limited environments by altering their metabolic strategies with more energy in producing enzymes related to nutrient acquisition, which results in excess carbon overflow for respiration (Manzoni et al. 2012; Geyer et al. 2016; Mehnaz et al. 2019). Soil CUE decreased significantly due to phosphorus limitation (Mganga et al.,

2022). High nutrient availability (simple SOM chemical structure and weak mineral-organic associations) is accompanied by high microbial CUE (Duan et al. 2023). Stoichiometric studies have shown that the C: N ratio can also significantly influence microbial CUE (Manzoni et al. 2012; Sinsabaugh et al. 2013; Takriti et al. 2018). At environments with a high C: N ratio (>10), microorganisms consume excess carbon through overflow respiration or producing other metabolites (such as proteins), thereby reducing CUE (Manzoni et al. 2012).

### Soil texture and aggregates

Soil texture has an impact on soil water effects, with coarser-textured soils having lower water content compared to finer-textured soils. In theory, Microorganisms in coarser-textured soils may be more prone to diffusion limitations associated with low water content, resulting lower CUE (Butcher et al. 2020). However, contrary to expectations, the content of clay was negatively correlated with the content of CUE (Oliver et al. 2021). Studies have found that sandy soils have higher microbial CUE and lower biomass turnover times compared to clay soils (Li et al. 2021; Pei et al. 2021). A reasonable explanation is that clay content strongly affects the diffusivity of the matrix and the accessibility of microorganisms (Krull et al. 2003). Substrate are stabilized by interactions with clay mineral surfaces. (Krull et al. 2003; Li et al. 2020a). It is mean that higher clay content can better bind with the substrate, reducing its diffusivity. Additionally, higher clay content also has reduced the accessibility of substrates to microorganisms by physically protection (Li et al. 2020b).

Soil aggregates directly affect microbial growth and activities by providing diverse environments for microorganisms (Hattori, 1988). In return, microorganisms are usually involved in the formation of aggregates by binding between particles (Lehmann, J. et al. 2015). The reason that macroaggregates have a higher CUE than microaggregates is that microaggregates have higher carbon stability, making them more difficult for microorganisms to utilize. (Bimüller et al. 2016; Najera et al. 2020). The SOM of microaggregates is formed by primary particles of plant and microbial debris combined with humus substances or polysaccharide polymers. Humus substances or polysaccharide polymers can better protect organic matter from decomposition (Kimura et al. 2012; Deneff et al. 2007). The low degradability of SOC in microaggregates reduces microbial mineralization (Tian et al. 2016), which may result in lower CUE. This conjecture has also been proved (Zhao et al. 2022; Li et al. 2024). Diversity in soil aggregate structure also imply complex nutrient supplies, leading to in organic matter degradability and microbial CUE (Mo et al. 2021). Macroaggregates usually represent an important site of nutrient (e.g., N,

P) accumulation (Green et al. 2005; Fonte et al. 2014). SOM has been proven to be more efficiently decomposed in N- and P-modified soils (Li et al., 2014). Therefore, compared with microaggregates, the SOM of macroaggregates appears to have higher degradability, leading to larger microbial CUE (Tian et al. 2016).

### Constraints of water and thermal conditions

The constraints of soil water and thermal conditions have significant impacts on soil microbial CUE, particularly in the context of climate warming and changes in soil moisture (Classen et al. 2015; Tian et al. 2023; Liu et al. 2024). Climate warming lead to an increase in temperature, temperature increases often trigger other chain reactions, and their impact on CUE is multifaceted. Higher temperature, on one hand, can lead to an increase in CUE and accelerate SOM decomposition. A five-year experiment found that warming trigger fundamental changes in the physiology of microbial communities in tropical forest soil, increasing CUE (Nottingham et al. 2019). On a global scale, Ye et al. (2019) incorporated microbial CUE and the relationship between mean annual temperature (MAT) and enzyme kinetics-MAT into a SOC model. By datasets of measured respiration (including 110 dryland soils distributed globally and two mainlands to global-scale cross-biome datasets), this work found a positive CUE-MAT relationship. In other words, microbial CUE tends to increase with increasing MAT, possibly because the availability of plant carbon inputs is generally higher in warmer climates (Bardgett et al. 2008).

On the other hand, some studies have found that CUE is resistant to warming, because of thermal adaptability of microorganisms. Simon et al. (2020) conducted measurements of soil microbial growth, CUE, and respiration under three distinct temperature and atmospheric CO<sub>2</sub> levels, as well as summer drought conditions. The authors found the individual and combined effects of climate change treatments on microbial growth and respiration rate were significant. But microbial CUE at the community level remains almost stable regardless of treatment or season. Similarly, study have found that the CUE of temperate forest soil in the 0–40 cm depth did not change significantly in response to warming (Spohn et al. 2016a). This situation is also found in a 3.3-year warming experiment with a 4 °C increase in the 0–100 cm depth of alpine grassland soil on the Qinghai-Tibet Plateau (Zhang et al. 2023b) or a 2-year warming experiment with a 4 °C increase in the 0–100 cm depth of tropical forest soil (Nottingham et al. 2020). The response of microbial CUE to warming also varies at different soil depths, which may be due to the difference in substrate structure and microbial community. For a short-term (110-day) experiment on soil warming in alpine grassland, it was revealed that nitrogen limitation induced by warming reduced

microbial CUE in the subsoil (30–40 cm), but not for the topsoil (0–10 cm) (Zhu et al. 2021). In another 4.5-year soil warming experiment with an increase of 4 °C, CUE was found to remain unchanged in the 0–60 cm depth, but decreased below 60 cm depth (Dove et al. 2021). In summary, the effect of climate warming on soil microbial CUE remains uncertain with complex environment of soil depth.

CUE is frequently reported to be reduced as the temperature increased (Frey et al. 2013; Rivkin and Legendre 2001; Wetterstedt and Agren 2011). The rate of maintenance respiration usually increases faster than the rate of new microbial biomass production under warming (Frey et al. 2013). Through a microbial-enzyme model to simulate the soil carbon response to a 5 °C temperature rise, it was found that CUE decreased under warming conditions (Allison et al. 2010). On a longer timescale, a 26-year soil warming experiment also revealed a decrease in CUE due to climate warming (Melillo et al. 2017). Shi et al. (2023) found through a 10-year forest soil warming experiment that soil warming led to increased non-biological adsorption of phosphorus. This condition lowered the availability of soil phosphorus, ultimately resulting in phosphorus limitation. To obtain the necessary phosphorus for growth, microorganisms increased the secretion of phosphatases, thereby reducing the carbon used for growth. Li et al. (2018) combined a microbial enzyme model with 22 years of carbon cycle measurements in Harvard Forest, using a probabilistic inversion method. They found that the increased temperature reduced CUE, and soil warming increased the temperature sensitivity of CUE. Notably, in an 11-year warming experiment, traditional tillage topsoil showed a 77% decrease in CUE due to warming, while CUE under conservation tillage increased by 29.1% (Wang et al. 2022). Microorganisms were regarded to effectively utilize richer and higher-quality substrates to build biomass during conservation tillage (Wang et al. 2022). In terms of most studies, the response of CUE to warming tends to decrease. However, considering the changes in microbial communities, the increase in carbon input, and the thermal adaptability of microorganisms that may be caused by warming, its specific response requires further analyzed.

Soil moisture regulates microbial activity through two primary physical mechanisms. The first one is the drying effect caused by low water potential. The decrease in extracellular water potential requires the accumulation of compatible solutes within microbial cells to balance the extracellular and intracellular water potentials (Rath and Rousk 2015; Rath et al. 2016). The second is the substrate diffusion limitation due to low water content (Herron et al. 2009). Substrate diffusion limitation is a physical constraint on soil microbial communities, limiting access of microbial communities to substrates (Weerts et al.

2001). But the influence of soil moisture on microbial CUE still needs to be specifically discussed. Study have found that drought conditions increased the fungal-to-bacterial ratio, thereby enhancing microbial CUE (Sun et al. 2022; Butcher et al. 2020). Similarly, in a short-term water stress event, CUE increases as osmoregulatory solutes and storage compounds are accumulated (Uhlrova, et al. 2005; Herron et al. 2009). However, in the long-term water stress event, CUE is reduced by repeated stress events, as the C costs for water stress responses become apparent (Tiemann and Billings 2011). Microorganisms also exhibit different responses under different humidity conditions, which further affect microbial CUE. Only under high humidity conditions, microbial species exhibit complementary effects by cross-feed shared resources (e.g., one species can utilize degradation products from neighboring species as substrates for growth). These effects enable more efficient growth, which may benefit microbial CUE (Domeignoz-Horta et al. 2020). In conclusion, when the interaction of microorganisms is not considered, short-term drought tends to reduce CUE, but the effect may be opposite under long-term drought.

#### External nutrient input

Most studies have found that under the condition of applying only N or adding N, P, and K simultaneously, CUE tends to increase (Poeplau et al. 2019; Zhang et al. 2022; Xu et al. 2024). The increase in CUE under N addition conditions can be attributed to several factors. First, it inhibits microbial respiration, which leads to an imbalance in the C: N ratio. This imbalance, in turn, enhances nitrogen utilization efficiency and reduces carbon investment in the production of extracellular enzymes for nitrogen acquisition. (Malik et al. 2020; Wang et al. 2023b; Yang et al. 2023). Second, some studies propose that the increase in CUE is not driven by stoichiometric imbalance. Instead, a decrease in the mineral protection of SOC increases the accessibility of microorganisms to carbon. (Feng et al. 2022). N induction led to changes in fungal composition, resulting in the production of more oxalic acid and causing changes in plant characteristics and community composition. Both of these aspects weakened mineral protection (Feng et al. 2022), organic matter is decomposed into smaller and more easily degradable molecular compounds, which are more conducive to microbial absorption and assimilation (Wallenstein and Weintraub 2008). Nitrogen-induced stress, such as soil acidification (Treseder 2008), may also trigger energy-intensive metabolic pathways or even directly cause microbial death (Jones et al. 2019; Horn et al. 2021), thus affecting microbial CUE.

Ma et al. (2023b) conducted a study examining the microbial CUE of soil layers after nitrogen fertilizer application. In contrast to previous findings, the authors

observed that both short-term (2 years) and long-term (10 years) nitrogen addition did not significantly affect microbial CUE. It attributed to the slight increase in carbon allocation to microbial biomass production, which was insufficient to significantly change CUE (Ma et al. 2023b). In a similar study, Jiang et al. (2024) found that nitrogen fertilizer application in tropical forests did not affect the CUE of surface soil (0–10 cm). However, the CUE in deep soil (60–80 cm) increased by 25.5%, likely due to the decreased ratio of fungi to bacteria and the C: N ratio. In a 6-year nitrogen addition experiment, it was also found that the decrease in the ratio of fungi to bacteria in temperate grasslands led to a decrease in CUE (Riggs and Hobbie 2016).

Long-term fertilization (N, P, K) conditions enhanced microbial CUE (Li et al. 2021). Adding nitrogen and phosphorus to the soil likely reduces metabolic costs for microorganisms to acquire nutrients. It also eliminates the need for additional enzymes and enables more carbon to be used for growth, ultimately enhancing CUE (Manzoni et al. 2012; Spohn et al. 2016b). Xu et al. (2024) found that the microbial CUE in soil treated with mineral fertilizer (N, P, K) alone increased by 22% compared to unfertilized soil. Additionally, the addition of Ca can also influence soil microbial CUE. Ca addition, on one hand, can promote bacteria production and affect the microbial community. On the other hand, it enhances the stability of microbial by-products by increasing the binding of litter and microbial by-products, resulting in a 45% increase in CUE (Shabtai et al. 2023). In general, if the input of external nutrients does not significantly change the ratio of fungi to bacteria in the soil, it tends to increase microbial CUE.

## Prospects

From a research perspective on soil microbial CUE, most current studies concentrate on single-site, single-factor impacts. These studies primarily examine changes in fungal and bacterial functional groups, carbon substrate quality, nutritional limitations, as well as the impacts of carbon, nitrogen, and phosphorus addition, organic fertilizer application, temperature rise, soil pH, moisture, aggregate size, and plant community composition and changes (Fig. 2). Given the complex interactions between soil, plants, and microorganisms, future research should pay more attention to the combined effects of multiple factors on soil microbial CUE.

In consideration of soil layers observed for soil microbial CUE, many studies concentrate primarily on the 0–15 (20) cm surface soil layer, overlooking the CUE at deeper soil depths. Moreover, further research should encompass other aspects, such as temperature sensitivity and substrate quality, across different soil depths. A comprehensive investigation is warranted to elucidate how

the soil microbial CUE varies with the multi-factorial changes across soil profile.

As regards the types of ecosystems observed for soil microbial CUE, present studies cover diverse types of ecosystems. Study places significant focus on the comparison and analysis among different ecosystems, including farmland, grassland, shrubland and forest but blue carbon habitats, also, deserve more attention. Currently, the time range of studies tends to be short-term. Additionally, in compared to local study, comprehensive focus on a global scale is still limited, but necessary.

## Acknowledgements

Not applicable.

## Author contributions

L.X. and G.H. conceptualized this study; R.D., J.L., L.X., E.L., L.Z., M.Z. have participated in writing the original draft of manuscript.

## Funding

This work was supported by National Natural Science Foundation of China (42077025, 42307437), Youth Innovation Promotion Association of CAS (2021213), Research Project of Dezhou University (2023XKZX004), Natural Science Foundation of Heilongjiang Province in China (YQ2023D007), and Natural Science Foundation of Jilin Province in China (20240101052JC).

## Data availability

Data sharing is not applicable to this article as no datasets were generated or analysed during the current study.

## Declarations

### Ethics approval and consent to participate

Not applicable.

### Consent for publication

Not applicable.

### Competing interests

Leilei Xiao is an Associate Editor for *Annals of Microbiology*. The authors declare that they have no other competing interests.

Received: 27 August 2024 / Accepted: 30 October 2024

Published online: 11 November 2024

## References

- Adingo S, Yu JR, Liu XL, Li XD, Jing S, Zhang X (2021) Variation of soil microbial carbon use efficiency (CUE) and its influence mechanism in the context of global environmental change: a review. *PeerJ* 9:e12131. <https://doi.org/10.717/peerj.12131>
- Agren GI, Bosatta N (1987) Theoretical analysis of the Long-Term Dynamics of Carbon and Nitrogen in Soils. *Ecology* 68:1181–1189. <https://doi.org/10.2307/1939202>
- Allison SD (2014) Modeling adaptation of carbon use efficiency in microbial communities. *Front Microbiol* 5. <https://doi.org/10.3389/fmicb.2014.00571>
- Allison SD, Wallenstein MD, Bradford MA (2010) Soil-carbon response to warming dependent on microbial physiology. *Nat Geosci* 3:336–340. <https://doi.org/10.1038/ngeo846>
- Alvarez R, Steinbach HS, De Paepe JL (2017) Cover crop effects on soils and subsequent crops in the pampas: a meta-analysis. *Soil till Res* 170:53–65. <https://doi.org/10.1016/j.still.2017.03.005>
- Anderson TH, Domsch KH (2010) Soil microbial biomass: the eco-physiological approach. *Soil Biol Biochem* 42:2039–2043. <https://doi.org/10.1016/j.soilbio.2010.06.026>

- Anthony MA, Celenza JL, Armstrong A, Frey SD (2020) Indolic glucosinolate pathway provides resistance to mycorrhizal fungal colonization in a non-host Brassicaceae. *Ecosphere* 11:e03100. <https://doi.org/10.1002/ecs2.3100>
- Bardgett RD, Freeman C, Ostle NJ (2008) Microbial contributions to climate change through carbon cycle feedbacks. *ISME J* 2:805–814. <https://doi.org/10.1038/ismej.2008.58>
- Bimüller C, Kreyling O, Kölbl A, Lützw M, Kögel-Knabner I (2016) Carbon and nitrogen mineralization in hierarchically structured aggregates of different size. *Soil till Res* 160:23–33. <https://doi.org/10.1016/j.still.2015.12.011>
- Butcher KR, Nasto MK, Norton JM, Stark JM (2020) Physical mechanisms for soil moisture effects on microbial carbon-use efficiency in a sandy loam soil in the western United States. *Soil Biol Biochem* 150:107969. <https://doi.org/10.1016/j.soilbio.2020.107969>
- Chen Y, Hu H (2024) Impacts of tree species diversity on microbial carbon use efficiency. *Global Change Biol* 30:e17015. <https://doi.org/10.1111/gcb.17015>
- Classen AT, Sundqvist MK, Henning JA, Newman GS, Moore JAM, Cregger MA, Moorhead LC, Patterson CM (2015) Direct and indirect effects of climate change on soil microbial and soil microbial-plant interactions: what lies ahead? *Ecosphere* 6:1–21. <https://doi.org/10.1890/ES15-00217.1>
- Craig ME, Geyer KM, Beidler KV, Brzostek ER, Frey SD, Stuart Grandy A, Liang C, Phillips RP (2022) Fast-decaying plant litter enhances soil carbon in temperate forests but not through microbial physiological traits. *Nat Commun* 13:1229. <https://doi.org/10.1038/s41467-022-28715-9>
- Denef K, Zotarelli L, Boddey RM, Six J (2007) Microaggregate-associated carbon as a diagnostic fraction for management-induced changes in soil organic carbon in two oxisols. *Soil Biol Biochem* 39:1165e1172
- Domeignoz-Horta LA, Pold G, Liu X-JA, Frey SD, Melillo JM, DeAngelis KM (2020) Microbial diversity drives carbon use efficiency in a model soil. *Nat Commun* 11:3684. <https://doi.org/10.1038/s41467-020-17502-z>
- Dove NC, Torn MS, Hart SC, Taş N (2021) Metabolic capabilities mute positive response to direct and indirect impacts of warming throughout the soil profile. *Nat Commun* 12:2089. <https://doi.org/10.1038/s41467-021-22408-5>
- Duan P, Fu R, Nottingham AT, Domeignoz-Horta LA, Yang X, Du H, Wang K, Li D (2023) Tree species diversity increases soil microbial carbon use efficiency in a subtropical forest. *Global Change Biol* 29:7131–7144. <https://doi.org/10.1111/gcb.16971>
- Feng X, Qin S, Zhang D, Chen P, Hu J, Wang G, Liu Y, Wei B, Li Q, Yang Y, Chen L (2022) Nitrogen input enhances microbial carbon use efficiency by altering plant–microbe–mineral interactions. *Global Change Biol* 28:4845–4860. <https://doi.org/10.1111/gcb.16229>
- Fonte SJ, Nesper M, Hegglin D, Velasquez JE, Ramirez B, Rao IM, Bernasconi SM, Bünemann EK, Frossard E, Oberson A (2014) Pasture degradation impacts soil phosphorus storage via changes to aggregate-associated soil organic matter in highly weathered tropical soils. *Soil Biol Biochem* 68:150e157. <https://doi.org/10.1016/j.soilbio.2013.09.025>
- Frey SD, Lee J, Melillo JM, Six J (2013) The temperature response of soil microbial efficiency and its feedback to climate. *Nat Clim Change* 3:395–398. <https://doi.org/10.1038/nclimate1796>
- García-Palacios P, Crowther TW, Dacal M, Hartley IP, Reinsch S, Rinnan R, Rousk J, Van Den Hoogen J, Ye JS, Bradford MA (2021) Evidence for large microbial-mediated losses of soil carbon under anthropogenic warming. *Nat Rev Earth Environ* 2:507–517. <https://doi.org/10.1038/s43017-021-00178-4>
- Geyer KM, Kyker-Snowman E, Grandy AS, Frey SD (2016) Microbial carbon use efficiency: accounting for population, community, and ecosystem-scale controls over the fate of metabolized organic matter. *Biogeochemistry* 127:173–188. <https://doi.org/10.1007/s10533-016-0191-y>
- Gommers PJF, Van Schie BJ, Van Dijken JP, Kuenen JG (1988) Biochemical limits to microbial growth yields: an analysis of mixed substrate utilization. *Biotechnol Bioeng* 32(1):6–94. <https://doi.org/10.1002/bit.260320112>
- Gottschall F, Cesarz S, Auge H, Kovach KR, Mori AS, Nock CA, Eisenhauer N (2022) Spatiotemporal dynamics of abiotic and biotic properties explain biodiversity–ecosystem-functioning relationships. *Ecol Monogr* 92:e01490. <https://doi.org/10.1002/ecm.1490>
- Green VS, Cavigelli MA, Dao TH, Flanagan DC (2005) Soil physical properties and aggregate-associated C, N and P distributions in organic and conventional cropping systems. *Soil Sci* 170:822e831. <https://doi.org/10.1097/01.ss.0000190509.18428.fe>
- Hartmann A, Schmid M, Tuinen DV, Berg G (2009) Plant-driven selection of microbes. *Plant Soil* 321:235–257. <https://doi.org/10.1007/s11104-008-9814-y>
- Herron PM, Stark JM, Holt C, Hooker T, Cardon ZG (2009) Microbial growth efficiencies across a soil moisture gradient assessed using 13 C-acetic acid vapor and 15 N-ammonia gas. *Soil Biol Biochem* 41:1262–1269. <https://doi.org/10.1016/j.soilbio.2009.03.010>
- Hervé V, Junier T, Bindschedler S, Verrecchia E, Junier P (2016) Diversity and ecology of oxalotrophic bacteria. *World J Microb Biot* 32(2):28. <https://doi.org/10.1007/s11274-015-1982-3>
- Horn EL, Cooledge EC, Jones DL, Hoyle FC, Brailsford FL, Murphy DV (2021) Addition of base cations increases microbial carbon use efficiency and biomass in acidic soils. *Soil Biol Biochem* 161:108392. <https://doi.org/10.1016/j.soilbio.2021.108392>
- Iven H, Walker TW, Anthony M (2023) Biotic interactions in soil are underestimated drivers of Microbial Carbon Use Efficiency. *Curr Microbiol* 80:13. <https://doi.org/10.1007/s00284-022-02979-2>
- Jiang Y, Su T, Wang H, Yang Q, Lu J, Fu Q, Mao H, Xu W, Luo Y, Liu W, Yang H, Fang M (2024) Deep soil microbial carbon use efficiency responds stronger to nitrogen deposition than top soil in tropical forests, southern China. *Plant Soil* 500:605–622. <https://doi.org/10.1007/s11104-024-06509-w>
- Jones DL, Cooledge EC, Hoyle FC, Griffiths RI, Murphy DV (2019) pH and exchangeable aluminum are major regulators of microbial energy flow and carbon use efficiency in soil microbial communities. *Soil Biol Biochem* 138:107584. <https://doi.org/10.1016/j.soilbio.2019.107584>
- Kimura SD, Melling L, Goh KJ (2012) Influence of soil aggregate size on greenhouse gas emission and uptake rate from tropical peat soil in forest and different oil palm development years. *Geoderma* 185:1e5. <https://doi.org/10.1016/j.geoderma.2012.03.026>
- Krull ES, Baldock JA, Skjemstad JO (2003) Importance of mechanisms and processes of the stabilisation of soil organic matter for modelling carbon turnover. *Funct Plant Biol* 30:207–222. <https://doi.org/10.1071/FP02085>
- Lange M, Eisenhauer N, Sierra CA, Bessler H, Engels C, Griffiths RI, Mellado-Vázquez PG, Malik AA, Roy J, Scheu S, Steinbeiss S, Thomson BC, Trumbore SE, Gleixner G (2015) Plant diversity increases soil microbial activity and soil carbon storage. *Nat Commun* 6:6707. <https://doi.org/10.1038/ncomms7707>
- Lehmann J, Kleber M (2015) The contentious nature of soil organic matter. *Nature* 528:60–68. <https://doi.org/10.1038/nature16069>
- Li J, Wang G, Mayes MA, Allison SD, Frey SD, Shi Z, Hu X, Luo Y, Melillo JM (2018) Reduced carbon use efficiency and increased microbial turnover with soil warming. *Global Change Biol* 25:900–910. <https://doi.org/10.1111/gcb.14517>
- Li J, Nie M, Pendall E (2020a) Soil physico-chemical properties are more important than microbial diversity and enzyme activity in controlling carbon and nitrogen stocks near Sydney, Australia. *Geoderma* 366:114201. <https://doi.org/10.1016/j.geoderma.2020.114201>
- Li J, Nie M, Powell JR et al (2020b) Soil physico-chemical properties are critical for predicting carbon storage and nutrient availability across Australia. *Environ Res Lett* 15:094088. <https://doi.org/10.1088/1748-9326/ab9f7e>
- Li J, Pei J, Dijkstra FA, Nie M, Pendall E (2021) Microbial carbon use efficiency, biomass residence time and temperature sensitivity across ecosystems and soil depths. *Soil Biol Biochem* 154:108117. <https://doi.org/10.1016/j.soilbio.2020.108117>
- Li Y, Zhang X, Li M, Liu J, Zhang K, Li Z (2024) Long-term straw mulching alleviates microbial nutrient limitations and increases carbon-use efficiency within aggregates. *Soil Use Manage* 40:e13058. <https://doi.org/10.1111/sum.13058>
- Liang C, Schimel JP, Jastrow JD (2017) The importance of anabolism in microbial control over soil carbon storage. *Nat Microbiol* 2:17105. <https://doi.org/10.1038/nmicrobiol.2017.105>
- Liang C, Amelung W, Lehmann J, Kästner M (2019) Quantitative assessment of microbial necromass contribution to soil organic matter. *Global Change Biol* 25:3578–3590. <https://doi.org/10.1111/gcb.14781>
- Liu X, Lai Q, Yin S, Bao Y, Tong S, Adiya Z, Sanjiv A, Gao R (2024) Spatio-temporal patterns and control mechanism of the ecosystem carbon use efficiency across the Mongolian Plateau. *Sci Total Environ* 907:167883. <https://doi.org/10.1016/j.scitotenv.2023.167883>
- Lu JZ, Scheu S (2021) Response of soil microbial communities to mixed beech-conifer forests varies with site conditions. *Soil Biol Biochem* 155:108155. <https://doi.org/10.1016/j.soilbio.2021.108155>
- Ma S, Zhu W, Wang WW, Li X, Sheng ZL (2023a) Microbial assemblies with distinct trophic strategies drive changes in soil microbial carbon use efficiency along vegetation primary succession in a glacier retreat area of the southeastern tibetan Plateau. *Sci Total Environ* 867:161587. <https://doi.org/10.1016/j.scitotenv.2023.161587>
- Ma S, Zhu W, Wang W, Li X, Sheng Z (2023b) Effects of short- and long-term nutrient addition on microbial carbon use efficiency and carbon accumulation efficiency in the tibetan alpine grassland. *Soil till Res* 229:105657. <https://doi.org/10.1016/j.still.2023.105657>

- Malik AA, Martiny JBH, Brodie EL, Martiny AC, Treseder KK, Allison SD (2020) Defining trait-based microbial strategies with consequences for soil carbon cycling under climate change. *ISME J* 14:1–9. <https://doi.org/10.1038/s41396-019-0510-0>
- Manzoni S, Porporato A (2009) Soil carbon and nitrogen mineralization: theory and models across scales. *Soil Biol Biochem* 41:1355–1379. <https://doi.org/10.1016/j.soilbio.2009.02.031>
- Manzoni S, Taylor P, Richter A, Porporato A, Agren GI (2012) Environmental and stoichiometric controls on microbial carbon-use efficiency in soils. *New Phytol* 196:79–91. <https://doi.org/10.1111/j.1469-8137.2012.04225.x>
- Mehnaz KR, Corneo PE, Keitel C, Dijkstra FA (2019) Carbon and phosphorus addition effects on microbial carbon use efficiency, soil organic matter priming, gross nitrogen mineralization and nitrous oxide emission from soil. *Soil Biol Biochem* 134:175–186. <https://doi.org/10.1016/j.soilbio.2019.04.003>
- Melillo JM, Frey SD, DeAngelis KM, Werner WJ, Bernard MJ, Bowles FP, Pold G, Knorr MA, Grandy AS (2017) Long-term pattern and magnitude of soil carbon feedback to the climate system in a warming world. *Science* 358:101–105. <https://doi.org/10.1126/science.aan2874>
- Mo F, Zhang YY, Liu Y, Liao YC (2021) Microbial carbon-use efficiency and straw-induced priming effect within soil aggregates are regulated by tillage history and balanced nutrient supply. *Biol Fertil Soils* 57:409–420. <https://doi.org/10.1007/s00374-021-01540-w>
- Mooshammer M, Wanek W, Zechmeister-Boltenstern S, Richter A (2014) Stoichiometric imbalances between terrestrial decomposer communities and their resources: mechanisms and implications of microbial adaptations to their resources. *Front Microbiol* 5. <https://doi.org/10.3389/fmicb.2014.00022>
- Moreau D, Pivato B, Bru D, Busset H, Deau F, Favre C, Matejcek A, Strbik F, Philippot L, Mougél C (2015) Plant traits related to nitrogen uptake influence plant-microbe competition. *Ecology* 96:2300–2310. <https://doi.org/10.1890/14-1761.1>
- Mori AS, Cornelissen JHC, Fujii S, Okada K, Isbell F (2020) A meta-analysis on decomposition quantifies afterlife effects of plant diversity as a global change driver. *Nat Commun* 11:4547. <https://doi.org/10.1038/s41467-020-18296-w>
- Najera F, Dippold MA, Boy J, Seguel O, Koester M, Stock S, Merino C, Kuzyakov Y, Matus F (2020) Effects of drying/rewetting on soil aggregate dynamics and implications for organic matter turnover. *Biol Fertil Soils* 56:893–905. <https://doi.org/10.1007/s00374-020-01469-6>
- Nottingham AT, Whitaker J, Ostle NJ, Bardgett RD, McNamara NP, Fierer N, Salinas N, Cahuana AJQ, Turner BL, Meir P (2019) Microbial responses to warming enhance soil carbon loss following translocation across a tropical forest elevation gradient. *Ecol Lett* 22:1889–1899. <https://doi.org/10.1111/ele.13379>
- Nottingham AT, Meir P, Velasquez E, Turner BL (2020) Soil carbon loss by experimental warming in a tropical forest. *Nature* 584:234–237. <https://doi.org/10.1038/s41586-020-2566-4>
- Oliver EE, Houlton BZ, Lipson DA (2021) Controls on soil microbial carbon use efficiency over long-term ecosystem development. *Biogeochemistry* 152:309–325. <https://doi.org/10.1007/s10533-021-00758-y>
- Pei J, Li J, Mia S et al (2021) Biochar aging increased microbial carbon use efficiency but decreased biomass turnover time. *Geoderma* 382:114710. <https://doi.org/10.1016/j.geoderma.2020.114710>
- Peng S, Chen HYH (2021) Global responses of fine root biomass and traits to plant species mixtures in terrestrial ecosystems. *Global Ecol Biogeogr* 30:289–304. <https://doi.org/10.1111/geb.13205>
- Poeplau C, Helfrich M, Dechow R et al (2019) Increased microbial anabolism contributes to soil carbon sequestration by mineral fertilization in temperate grasslands. *Soil Biol Biochem* 130:167–176. <https://doi.org/10.1016/j.soilbio.2018.12.019>
- Powell JR, Bennett AE (2016) Unpredictable assembly of arbuscular mycorrhizal fungal communities. *Pedobiologia* 59:11–15. <https://doi.org/10.1016/j.pedobi.2014.11.001>
- Prommer J, Walker TWN, Wanek W, Braun J, Zezula D, Hu Y, Hofhansl F, Richter A (2020) Increased microbial growth, biomass, and turnover drive soil organic carbon accumulation at higher plant diversity. *Global Change Biol* 26:669–681. <https://doi.org/10.1111/gcb.14777>
- Rath KM, Rousk J (2015) Salt effects on the soil microbial decomposer community and their role in organic carbon cycling: a review. *Soil Biol Biochem* 81:108–123. <https://doi.org/10.1016/j.soilbio.2014.11.001>
- Rath KM, Maheshwari A, Bengtson P, Rousk J (2016) Comparative toxicities of salts on microbial processes in Soil. *Appl Environ Microbiol* 82:2012–2020. <https://doi.org/10.1128/AEM.04052-15>
- Riggs CE, Hobbie SE (2016) Mechanisms driving the soil organic matter decomposition response to nitrogen enrichment in grassland soils. *Soil Biol Biochem* 99:54–65. <https://doi.org/10.1016/j.soilbio.2016.04.023>
- Rivkin RB, Legendre L (2001) Biogenic carbon cycling in the upper ocean: effects of microbial respiration. *Science* 291:2398–2400. <https://doi.org/10.1126/science.291.5512.2398>
- Roels JA (1980) Application of macroscopic principles to microbial metabolism. *Biotechnol Bioeng* 22(12):2457–2514. <https://doi.org/10.1002/bit.260221202>
- Schroeder J, Dämmtircá C, Bölscher T, Chenu C, Elsgaard L, Tebbe CC, Skadell L, Poeplau C (2024) Liming effects on microbial carbon use efficiency and its potential consequences for soil organic carbon stocks. *Soil Biol Biochem* 191:109342. <https://doi.org/10.1016/j.soilbio.2024.109342>
- Shabtai IA, Wilhelm RC, Schweizer SA, Höschel C, Buckley DH, Lehmann J (2023) Calcium promotes persistent soil organic matter by altering microbial transformation of plant litter. *Nat Commun* 14:6609. <https://doi.org/10.1038/s41467-023-42291-6>
- Shi C, Urbina-Malo C, Tian Y, Heinzle J, Kwatcho Kengdo S, Inselsbacher E, Borken W, Schindlbacher A, Wanek W (2023) Does long-term soil warming affect microbial element limitation? A test by short-term assays of microbial growth responses to labile C, N and P additions. *Global Change Biol* 29:2188–2202. <https://doi.org/10.1111/gcb.16591>
- Silva-Sánchez A, Soares M, Rousk J (2019) Testing the dependence of microbial growth and carbon use efficiency on nitrogen availability, pH, and organic matter quality. *Soil Biol Biochem* 134:25–35. <https://doi.org/10.1016/j.soilbio.2019.03.008>
- Simon E, Canarini A, Martin V, Séneca J, Böckle T, Reinthaler D, Pötsch EM, Piepho HP, Bahn M, Wanek W, Richter A (2020) Microbial growth and carbon use efficiency show seasonal responses in a multifactorial climate change experiment. *Commun Biol* 3:584. <https://doi.org/10.1038/s42003-020-01317-1>
- Sinsabaugh RL, Manzoni S, Moorhead DL, Richter A (2013) Carbon use efficiency of microbial communities: stoichiometry, methodology and modelling. *Ecol Lett* 16:930–939. <https://doi.org/10.1111/ele.12113>
- Sinsabaugh RL, Turner BL, Talbot JM, Waring BG, Powers JS, Kuske CR, Moorhead DL, Follstad Shah JJ (2016) Stoichiometry of microbial carbon use efficiency in soils. *Ecol Monogr* 86:172–189. <https://doi.org/10.1890/15-2110.1>
- Sokol NW, Slessarev E, Marschmann GL, Nicolas A, Blazewicz SJ, Brodie EL, Firestone MK, Foley MM, Hestrin R, Hungate BA, Koch BJ, Stone BW, Sullivan MB, Zablocki O, Pett-Ridge J (2022) Life and death in the soil microbiome: how ecological processes influence biogeochemistry. *Nat Rev Microbiol* 20:415–430. <https://doi.org/10.1038/s41579-022-00695-z>
- Spohn M, Klaus K, Wanek W, Richter A (2016a) Microbial carbon use efficiency and biomass turnover times depending on soil depth – implications for carbon cycling. *Soil Biol Biochem* 96:74–81. <https://doi.org/10.1016/j.soilbio.2016.01.016>
- Spohn M, Pötsch EM, Eichorst SA, Woeken D, Wanek W, Richter A (2016b) Soil microbial carbon use efficiency and biomass turnover in a long-term fertilization experiment in a temperate grassland. *Soil Biol Biochem* 97:168–175. <https://doi.org/10.1016/j.soilbio.2016.03.008>
- Sun Y, Wang C, Ruan H (2022) Increased microbial carbon and nitrogen use efficiencies under drought stress in a poplar plantation. *For Ecol Manag* 519:120341. <https://doi.org/10.1016/j.foreco.2022.120341>
- Takriti M, Wild B, Schnecker J, Mooshammer M, Knoltsch A, Lashchinskiy N, Eloy Alves RJ, Gentsch N, Gittel A, Mikutta R, Wanek W, Richter A (2018) Soil organic matter quality exerts a stronger control than stoichiometry on microbial substrate use efficiency along a latitudinal transect. *Soil Biol Biochem* 121:212–220. <https://doi.org/10.1016/j.soilbio.2018.02.022>
- Tao F, Huang Y, Hungate BA, Manzoni S, Frey SD, Schmidt MW, Reichstein M, Carvalhais N, Clais P, Jiang L, Lehmann J, Wang YP, Houlton BZ, Ahrens B, Mishra U, Hugelius G, Hocking TD, Lu X, Shi Z, Viatkin K, Vargas R, Yigini Y, Omuto C, Malik AA, Peralta G, Cuevas-Corona R, Di Paolo LE, Luotto I, Liao C, Liang YS, Saynes VS, Huang X, Luo Y (2023) Microbial carbon use efficiency promotes global soil carbon storage. *Nature* 618:981–985. <https://doi.org/10.1038/s41586-023-06042-3>
- Tian J, Pausch J, Yu G, Blagodatskaya E, Kuzyakov Y (2016) Aggregate size and glucose level affect priming sources: a three-source-partitioning study. *Soil Biol Biochem* 97:199–210. <https://doi.org/10.1016/j.soilbio.2016.03.013>
- Tian Y, Schindlbacher A, Malo CU, Shi C, Heinzle J, Kwatcho Kengdo S, Inselsbacher E, Borken W, Wanek W (2023) Long-term warming of a forest soil reduces microbial biomass and its carbon and nitrogen use efficiencies. *Soil Biol Biochem* 184:109109. <https://doi.org/10.1016/j.soilbio.2023.109109>
- Tiemann LK, Billings SA (2011) Changes in variability of soil moisture alter microbial community C and N resource use. *Soil Biol Biochem* 43:1837–1847. <https://doi.org/10.1016/j.soilbio.2011.04.020>

- Treseder KK (2008) Nitrogen additions and microbial biomass: a meta-analysis of ecosystem studies. *Ecol Lett* 11:1111–1120. <https://doi.org/10.1111/j.1461-0248.2008.01230.x>
- von Stockar U, van der Wielen LAM (2013) *Biothermodynamics: the role of thermodynamics in biochemical Engineering*, 1st edn. EPFL, New York. <https://doi.org/10.1201/b15428>
- Wallenstein MD, Weintraub MN (2008) Emerging tools for measuring and modeling the in situ activity of soil extracellular enzymes. *Soil Biol Biochem* 40:2098–2106. <https://doi.org/10.1016/j.soilbio.2008.01.024>
- Wang M, Dungait JAJ, Wei X, Ge T, Hou R, Ouyang Z, Zhang F, Tian J (2022) Long-term warming increased microbial carbon use efficiency and turnover rate under conservation tillage system. *Soil Biol Biochem* 172:108770. <https://doi.org/10.1016/j.soilbio.2022.108770>
- Wang W, Zhu W, Li X, Ma S (2023b) Long-term nitrogen addition increased soil microbial carbon use efficiency in subalpine forests on the eastern edge of the Qinghai–Tibet Plateau. *Plant Soil* 482:553–565. <https://doi.org/10.1007/s11104-022-05710-z>
- Wang B, Hu W, Xue J, Jing Y, Zhu H, Ding H (2023a) Revealing the globally multi-scale controls of environmental factors on carbon use efficiency. *Sci Total Environ* 892:164634. <https://doi.org/10.1016/j.scitotenv.2023.164634>
- Watson BS, Bedair MF, Urbanczyk-Wochniak E, Huhman DV, Yang DS, Allen SN, Li W, Tang Y, Sumner LW (2015) Integrated Metabolomics and transcriptomics reveal enhanced Specialized Metabolism in *Medicago truncatula* Root Border cells. *Plant Physiol* 167:1699–1716. <https://doi.org/10.1104/pp.114.253054>
- Weerts AH, Kandhai D, Bouten W, Sloot PMA (2001) Tortuosity of an Unsaturated Sandy Soil Estimated using gas diffusion and bulk Soil Electrical Conductivity: comparing analogy-based models and Lattice–Boltzmann simulations. *Soil Sci Soc Amer J* 65:1577–1584. <https://doi.org/10.2136/sssaj2001.1577>
- Wetterstedt JAM, Agren GI (2011) Quality or decomposer efficiency -which is most important in the temperature response of litter decomposition? A modelling study using the GLUE methodology. *Biogeosciences* 8:477–487. <https://doi.org/10.1890/04-1254>
- Wu X, Zhang Z, Zhang J, Liu Y, Luo W, Mou G, Huang X (2023) Multiple factors jointly lead to the Lower Soil Microbial Carbon Use Efficiency of *Abies fanjingshanensis* in a typical Subtropical Forest in Southwest China. *Forests* 14:1716. <https://doi.org/10.3390/f14091716>
- Wu G, Su Y, Wang J, Lin S, Huang Z, Huang G (2024) Elevational patterns of microbial carbon use efficiency in a subtropical mountain forest. *Biol Fertil Soils* 60:5–15. <https://doi.org/10.1007/s00374-022-01694-1>
- Xiong J, Wang G, Richter A, DeLuca TH, Zhang W, Sun H, Hu Z, Sun X, Sun S (2023) Soil organic carbon accumulation and microbial carbon use efficiency in subalpine coniferous forest as influenced by forest floor vegetative communities. *Geoderma* 438:116648. <https://doi.org/10.1016/j.geoderma.2023.116648>
- Xu Q, Li L, Guo J, Guo H, Liu M, Guo S, Kuzyakov Y, Ling N, Shen Q (2024) Active microbial population dynamics and life strategies drive the enhanced carbon use efficiency in high-organic matter soils. *mBio* 15:e00177–e00124. <https://doi.org/10.1128/mbio.00177-24>
- Yang X, Duan P, Hicks L, Wang K, Li D (2023) Mechanisms underlying the responses of microbial carbon and nitrogen use efficiencies to nitrogen addition are mediated by topography in a subtropical forest. *Sci Total Environ* 880:163236. <https://doi.org/10.1016/j.scitotenv.2023.163236>
- Ye J, Bradford MA, Dacal M, Maestre FT, García-Palacios P (2019) Increasing microbial carbon use efficiency with warming predicts soil heterotrophic respiration globally. *Global Change Biol* 25:3354–3364. <https://doi.org/10.1111/gcb.14738>
- Zhang M, Li S, Wu X, Zheng F, Song X, Lu J, Liu X, Wang B, Abdelrhmana AA, Degré A (2022) Nitrogen addition mediates the effect of soil microbial diversity on microbial carbon use efficiency under long-term tillage practices. *Land Degrad Dev* 33:2258–2275. <https://doi.org/10.1002/ldr.4279>
- Zhang HY, Bao WK, Hu B, Hu H (2023a) Effect of vegetation type change on soil microbial carbon use efficiency: a review. *Acta Ecol Sin* 46(16):6878–6888. <https://doi.org/10.5846/stxb202204161016>
- Zhang Q, Qin W, Feng J, Li X, Zhang Z, He JS, Schimel JP, Zhu B (2023b) Whole-soil-profile warming does not change microbial carbon use efficiency in surface and deep soils. *Proc Natl Acad Sci USA* 120:e2302190120. <https://doi.org/10.1073/pnas.2302190120>
- Zhao X, Lu X, Yang J, Zhang D, Ren C, Wang X, Zhang X, Deng J (2022) Effects of Nitrogen Addition on Microbial Carbon Use Efficiency of Soil aggregates in Abandoned Grassland on the Loess Plateau of China. *Forests* 13:276. <https://doi.org/10.3390/f13020276>
- Zhu E, Cao ZJ, Jia J, Liu CZ, Zhang ZH, Wang H, Dai GH, He JS, Feng XJ (2021) Inactive and inefficient: warming and drought effect on microbial carbon processing in alpine grassland at depth. *Global Change Biol* 27:2241–2253. <https://doi.org/10.1111/gcb.15541>

## Publisher's note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.